Version with Markings to Show Changes Made

The amended paragraphs indicate deletions by ~~strikeout~~ and insertions by underlining.

In the specification, page 3, second full paragraph.

> *Ballabio et al.* (1991), disclose a ~~single-tube multiplex allele-specific~~ single-tube, multiplex allele-specific PCR test using two different dye-tagged fluorescent primers for detection of the ▲F508 cystic fibrosis mutation.

In the specification, page 11, first full paragraph.

> The primers must also be designed so that the size of the resulting amplification products differ in length, thereby facilitating assignment of alleles to individual loci during detection. Inappropriate selection of primers can produce several undesirable effects such as lack of amplification, amplification at multiple sites, primer dimer formation, undesirable interaction of primer sequences from different loci, production of alleles from one locus which overlap with alleles from another, or ~~requirement~~ the need for amplification conditions or protocols for the different loci which are incompatible in a multiplex. The synthesis of the primers is conducted by procedures known to those skilled in the art.

In the specification, page 18, third full paragraph.

> In this example, a DNA template was amplified at the individual loci HUMCSF1PO, HUMTPOX, HUMTH01, and HUMVWFA31 simultaneously in a single reaction vessel. The PCR amplifications were performed in 25µl volumes using 25ng template, 0.04U *Taq* DNA Polymerase/µl, 1x STR Buffer (50mM KCl, 10mM Tris-HCl (pH 9.0 at 25°C), 0.1% Triton X-100, 1.5mM $MgCl_2$ and 200µM each of dATP, dCTP, dGTP and dTTP), and using a Thermal Cycler 480 (Perkin Elmer Cetus). Amplification protocol 1, as described in Example 1, was employed. Eight

amplification primers were used in combination, including 1μM each HUMCSF1PO primer 2 ~~[SEQ. ID. 5]~~ [SEQ. ID. 6] and fluorescein-labeled primer 1 [SEQ. ID. 5], 0.15μM each HUMTPOX primer 1 [SEQ. ID. 29] and fluorescein-labeled primer 2 [SEQ. ID. 30], 0.2μM each HUMTH01 primer 2 [SEQ. ID. 28] and fluorescein-labeled primer 1 [SEQ. ID. 27], and 1μM each HUMVWFA31 primer 1 [SEQ. ID. 31] and fluorescein-labeled primer 2 [SEQ. ID. 32].